

An alternative kinetic evaluation of a reversible enzyme reaction

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One of the most investigated biochemical pathways is the glycolysis. While searching in literature databases, more than 35.000 publications can be found. Some of these publications studied the thermodynamics of the glycolysis and have postulated that this pathway is not thermodynamically feasible [1-4]. A distributed bottleneck was identified between reaction 5 and 6 but there are also some local bottlenecks with a single unfeasible or undetermined reaction [2, 3]. One example for such a reaction is reaction 9 of the glycolysis where the conversion from 2-phosphoglycerate to phosphoenolpyruvate takes place which is not irreversible but an equilibrium reaction. Thermodynamic and kinetic investigations of this reaction were done previously but just with standard instead of cell mimicking conditions. In addition to taking these conditions into account new thermodynamic research of this reaction should also include a kinetic analysis meaning that an equilibrium reaction cannot be interpreted using the usual Michaelis-Menten kinetics. The backward reaction from the product to enzyme-substrate-complex is neglected by this classical approach. Strangely enough, the most kinetic studies on reaction 9 of the glycolysis evaluate the data using the Michaelis-Menten model and therefore disregard the reversibility of the reaction [5-9]. There are some models which incorporate the reaction equilibrium, like the Hoh and Cord-Ruwisch approach [10] or the reversible Michaelis-Menten mechanism [11-17]. The reversible Michaelis-Menten mechanism results in a bad parameter fit due to of the high number of adjustable parameters [15, 18]. Therefore, we want to present an alternative way for data analysis of reversible reactions that combines a kinetic approach with irreversible thermodynamics and drastically reduces the number of required parameters. In addition to this fundamental result, we were able to describe quantitatively the influences of the different cell mimicking conditions on the kinetics of the reaction. The imitation of the crowding inside cells decreases the kinetics the most. The temperature dependency of our new kinetic constants follow the well-known Arrhenius relation. The dependency on the pH was also strong. Magnesium and ionic strength showed a weaker dependency. In the further course of the study, we want to apply our model to other reversible reactions of glycolysis and explore the predictive potential of thermodynamics for systems biology.

Keywords isothermal titration calorimetry; irreversible thermodynamics; glycolysis; reversible reaction

References

1. Li, X., et al., *A Database of Thermodynamic Quantities for the Reactions of Glycolysis and the Tricarboxylic Acid Cycle*. Journal of Physical Chemistry B, 2010. **114**(49): p. 16068-16082.
2. Maskow, T. and U. von Stockar, *How reliable are thermodynamic feasibility statements of biochemical pathways?* Biotechnology and Bioengineering, 2005. **92**(2): p. 223-230.
3. Vojinovic, V. and U. von Stockar, *Influence of Uncertainties in pH, pMg, Activity Coefficients, Metabolite Concentrations, and Other Factors on the Analysis of the Thermodynamic Feasibility of Metabolic Pathways*. Biotechnology and Bioengineering, 2009. **103**(4): p. 780-795.
4. Rajendran, T.E. and T. Muthukumarasamy, *Thermodynamic calculations of biochemical reaction systems at specified pH, pMg, and change in binding of hydrogen and magnesium ions*. Asia-Pacific Journal of Chemical Engineering, 2018. **13**(4): p. 15.
5. Zadvornyy, O.A., et al., *Biochemical and Structural Characterization of Enolase from Chloroflexus aurantiacus: Evidence for a Thermophilic Origin*. Front Bioeng Biotechnol, 2015. **3**: p. 74.
6. Pancholi, V. and V.A. Fischetti, *alpha-enolase, a novel strong plasmin(ogen) binding protein on the surface of pathogenic streptococci*. Journal of Biological Chemistry, 1998. **273**(23): p. 14503-14515.
7. Wold, F. and C.E. Ballou, *Studies on the enzyme enolase II. kinetic studies*. Journal of Biological Chemistry, 1957. **227**(1): p. 313-328.
8. Westhead, E.W. and B.G. Malmstrom, *The chemical kinetics of the enolase reaction with special reference to the use of mixed solvents*. Journal of Biological Chemistry, 1957. **228**(2): p. 655-671.
9. Kornblatt, M.J. and R. Musil, *The Inhibition of Yeast Enolase by Li⁺ and Na⁺*. Archives of Biochemistry and Biophysics, 1990. **277**(2): p. 301-305.
10. Hoh, C.Y. and R. CordRuwisch, *A practical kinetic model that considers endproduct inhibition in anaerobic digestion processes by including the equilibrium constant*. Biotechnology and Bioengineering, 1996. **51**(5): p. 597-604.
11. Keleti, T., *two rules of enzyme kinetics for reversible Michaelis-Menten mechanisms*. Febs Letters, 1986. **208**(1): p. 109-112.
12. Imperial, S.a.C., J.J., *Enzyme Kinetic Equations of Irreversible and Reversible Reactions in Metabolism*. Journal of Biosciences and Medicines, 2014. **2**: p. 24-29.
13. Miller, W.G. and R.A. Alberty, *Kinetics of the Reversible Michaelis-Menten Mechanism and the Applicability of the Steady-state Approximation*. Journal of the American Chemical Society, 1958. **80**(19): p. 5146-5151.
14. Paul, S. and G. Gangopadhyay, *Power law kinetics in reversible enzyme-catalyzed reaction due to diffusion*. Journal of Chemical Physics, 2003. **119**(6): p. 3501-3508.
15. Lee, L.W., et al., *Generic enzymatic rate equation under living conditions*. Journal of Biological Systems, 2007. **15**(4): p. 495-514.
16. Smith, W.G., *In Vivo Kinetics and the Reversible Michaelis-Menten Model*. Journal of Chemical Education, 1992. **69**(12): p. 981-984.
17. Brooks, S.P.J. and K.B. Storey, *A kinetic description of sequential, reversible, Michaelis-Menten reactions: practical application of theory to metabolic pathways*. Molecular and Cellular Biochemistry, 1992. **115**(1): p. 43-48.
18. Cha, S., *A Simple Method for Derivation of Rate Equations for Enzyme-catalyzed Reactions under the Rapid Equilibrium Assumption or Combined Assumptions of Equilibrium and Steady State* Journal of Biological Chemistry, 1968. **243**(4): p. 820-&.